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U.S. APPLICATION NO. (If known, see 37 CFR 1.5)

INTERNATIONAL APPLICATION NO.
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PRIORITY DATE CLAIMED
30 April 1999

TITLE OF INVENTION: DRY MICROORGANISM CULTURES AND THEIR PREPARATION

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Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. /X/ This is a FIRST submission of items concerning a filing under 35 U.S.C. 371.
 2. / / This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371.
 3. /X/ This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
 4. /x / A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
 5. /X/ A copy of the International Application as filed (35 U.S.C. 371(c)(2)).
 - a. /X/ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. / / has been transmitted by the International Bureau.
 - c. / / is not required, as the application was filed in the United States Receiving Office (RO/USO).
 6. /X/ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
 7. /X / Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)).
 - a. /X / are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. / / have been transmitted by the International Bureau.
 - c. / / have not been made; however, the time limit for making such amendments has NOT expired.
 - d. / / have not been made and will not be made.
 8. /X / A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
 9. /X / An oath or declaration of the inventor(s) (35 U.S.C. 171(c)(4)).
 10. / / A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).
- Items 11. to 16. below concern other document(s) or information included:
11. / / An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
 12. /X / An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
 13. /X / A FIRST preliminary amendment.
/ / A SECOND or SUBSEQUENT preliminary amendment.
 14. / / A substitute specification.
 15. / / A change of power of attorney and/or address letter.
 16. /x / Other items or information.
International Search Report
International Preliminary Examination Report

U.S. Appl. No. (If known) **09/673136** INTERNATIONAL APPLN. NO. PCT/EP99/02925ATTORNEY'S DOCKET NO.
48996

17. ☒ The following fees are submitted
 BASIC NATIONAL FEE (37 CFR 1.492(a)(1)-(5)):
 Search Report has been prepared by the
 EPO or JPO.....\$860.00

CALCULATIONS**PTO USE ONLY**

International preliminary examination fee paid to USPTO
 (37 CFR 1.482).....\$750.00

No international preliminary examination fee paid to
 USPTO (37 CFR 1.482) but international search fee paid
 to USPTO (37 CFR 1.445(a)(2)).....\$700.00

Neither international preliminary examination fee
 (37 CFR 1.482) nor international search fee
 (37 CFR 1.445(a)(2)) paid to USPTO\$ 970.00

International preliminary examination fee paid to
 USPTO (37 CFR 1.482) and all claims satisfied pro
 visions of PCT Article 33(2)-(4).....\$96.00

ENTER APPROPRIATE BASIC FEE AMOUNT = \$ 860.00

Surcharge of \$130.00 for furnishing the oath or declaration
 later than 1 / 20 / 30 months from the earliest
 claimed priority date (37 CFR 1.492(e)).

Claims	Number Filed	Number Extra	Rate
Total Claims	19	-20	X\$18.
Indep. Claims	3	-3	X\$80.
Multiple dependent claim(s) (if applicable)		+270.	
TOTAL OF ABOVE CALCULATION			= 860.00

Reduction of 1/2 for filing by small entity, if applicable.
 Verified Small Entity statement must also be filed
 (Note 37 CFR 1.9, 1.27, 1.28).

SUBTOTAL = 860.00

Processing fee of \$130. for furnishing the English
 translation later than 1 / 20 / 30 months from the
 earliest claimed priority date (37 CFR 1.492(f)).

TOTAL NATIONAL FEE = 860.00

Fee for recording the enclosed assignment (37 CFR 1.21(h)).
 The assignment must be accompanied by an appropriate cover
 sheet (37 CFR 3.28, 3.31) \$40.00 per property

TOTAL FEES ENCLOSED = \$ 900.00

Amount to be
 refunded: \$
 Charged \$

a. ☒ A check in the amount of \$ 900. to cover the above fees is enclosed.

b. ☐ Please charge my Deposit Account No. _____ in the amount of \$ _____ to cover the above fees. A duplicate copy of this sheet is enclosed.

c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. **11-0345**. A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

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OFFICE OF THE COMMISSIONER OF PATENTS AND TRADEMARKS
 U.S. DEPARTMENT OF COMMERCE

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BOX PCT

Filed: April 29, 1999

PRELIMINARY AMENDMENT

Sir:

Prior to examination, kindly amend the above-identified application as follows:

IN THE CLAIMS

Cancel claims 1-18 and insert new claims 19-37 on the attached sheets.

REMARKS

The claims in the case are claims 19-37. The claims were amended in the preliminary examination. The claims have been amended further to eliminate multiple dependency and to put it in better form for U.S. filing. No new matter is included.

Favorable action is solicited.

Respectfully submitted,

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--19. A dry microorganism culture which comprises at least one microorganism species in carrier-bound form, wherein the culture is present in the form of particles which

- a) have a particle size of at least about 0.1 mm and
- b) comprise from about 10^8 to 10^{12} cfu/g of at least one microorganism species; and
- c) are compressed.

20. A microorganism culture as claimed in claim 19, wherein the particles have been compressed under the action of a linear force from about 5 to 15 kN/cm or a pressure from about 90 to 160 MPa.

21. A microorganism culture as claimed in claim 19, wherein the compressed particles comprise compacted broken material having a diameter of from about 0.1 mm to about 2 mm.

22. A microorganism culture as claimed in claim 19, wherein the compressed particles comprise tablets having a diameter of from about 2 to 50 mm and a ratio of diameter to thickness of from about 1:0.1 to about 10:1.

23. A microorganism culture as claimed in claim 19, wherein it comprises, a further component, an effervescent additive.

24. A microorganism culture as claimed in claim 19, wherein, as carrier, it comprises at least one matrix material for embedding the microorganism cells with or without at least one further cell-stabilizing additive.

25. A microorganism culture as claimed in claim 19, wherein it comprises at least one lactic-acid-producing bacterial species.

26. A microorganism culture as claimed in claim 25, wherein the bacterial

species is selected from bacteria of the genus *Lactobacillus* sp.

27. A process for producing a dry microorganism culture, comprising at least one microorganism species in carrier-bound form, which comprises

- a) dissolving or suspending at least one substance suitable for forming a carrier in a liquid comprising at least one microorganism species,
- b) drying the resultant mixture in a spray-dryer, for the spray-drying use being made of a conditioned dried gas having a dew point of less than about +50°C, heated to a temperature in the range of above about 80°C, and
- c) removing the dried material from the spray dryer, this dried material having an exit temperature of from about 45 to 75°C.

28. A process as claimed in claim 27, wherein, in a further stage d), the dry material is subjected to a further drying at a temperature in the range from about 15 to 50°C in a gas atmosphere or in vacuo and/or at least one desiccant is added.

29. A process as claimed in claim 27, wherein, as dry material, a powder concentrate having a content of viable microorganisms of from about $5 \cdot 10^8$ to $1 \cdot 10^{12}$ cfu/g is obtained.

30. Dry compressed microorganism culture according to claim 19, obtained from a powder concentrate of microorganism culture dried in a spray-dryer, for the spray-drying use being made of a conditioned dried gas having a dew point of less than about +50°C, heated to a temperature in the range of above about 80°C.

31. A process for preparing a dry microorganism culture as claimed in claim 19, which comprises

- i) producing a powder concentrate of the microorganism culture by carrier-bound

- an? .
- spray-drying, carrier-bound freeze-drying or carrier-bound fluidized-bed drying,
 - ii) with or without admixing the powder concentrate with one or more coformulants and
 - iii) compacting or tableting this mixture.

32. A process as claimed in claim 31, wherein the compacted powder concentrate from stage iii) is broken, with or without classification.

33. A process for preparing a dry agglomerated microorganism culture, which comprises

- i) preparing a powder concentrate of the microorganism culture by carrier-bound spray-drying, carrier-bound freeze drying or carrier-bound fluidized-bed drying,
- ii) with our without admixing the powder concentrate with one or more coformulants and
- iii) agglomerating this mixture.

34. A process as claimed in claim 31, wherein the spray-drying is performed in a spray-dryer in which a conditioned dried gas is employed having a dew point of less than about +50°C, heated to a temperature in the range of above about 80°C.

35. A starter culture for foodstuffs and feedstuffs comprising a microorganism culture as claimed in claim 19, or prepared by a process for producing a dry microorganism culture, comprising at least one microorganism species in carrier-bound form, which comprises

- a) dissolving or suspending at least one substance suitable for forming a carrier in a liquid comprising at least one microorganism species,
- b) drying the resultant mixture in a spray-dryer, for the spray-drying use being

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Dry microorganism cultures and their preparation

- 5 The present invention relates to novel dry microorganism cultures which can be used in particular to prepare foodstuffs and feedstuffs, and to processes for preparing dry microorganism cultures.
- 10 A main area of application of microorganisms, such as bacteria and yeasts, is preparing foodstuffs and feedstuffs. Thus, for example, lactic acid bacteria, such as those of the genus *Streptococcus* sp. or *Lactobacillus* sp. are used in the preparation of milk products, such as sour cream, buttermilk,
- 15 yogurt, kefir, coumis, curd cheese and in the preparation of sourdough and for preserving uncooked sausage, such as salami. Lactic acid bacteria, such as those of the genus *Lactobacillus* sp., for instance, are also used in the production of feeds, such as silage.
- 20 The microorganism preparations required for preparing foodstuffs and feedstuffs are usually used in the form of starter cultures. These are generally not freshly prepared liquid cultures, but either cultures usually frozen in liquid nitrogen or dry
- 25 preparations. Dry preparations are usually preferred, since their transport and storage is technically less complex in comparison with frozen preparations.
- 30 Very varied types of dry preparations of microorganism cultures are known from the prior art. Thus, for example, EP-A-0 131 114 describes a *Lactobacillus* preparation where a bacterial cell suspension is applied to a pulverulent or granulated carrier composition and dried. However, to store the preparation it is necessary to package this in an oxygen-free protective gas
- 35 atmosphere. DD 840493952 proposes freeze-drying strains of cultured microorganisms for producing starter cultures, packaging them in film and storing them at 279 to 288 Kelvin. JP-A-06/217713 describes the production of special *Lactobacillus* starter cultures by spray-drying. EP-A-0 202 409 proposes
- 40 subjecting dry cultures to a wet granulation, processing the granules to form spherical particles and then drying them. In addition, proposals are made in a number of publications to provide coated dry bacterial preparations (cf. US-A-3,677,897 for example).
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A number of different processes are described in the prior art to produce dry microorganism preparations. In addition to the freeze-drying and fluidized-bed drying processes mentioned above, another alternative production method is spray-drying a microorganism suspension. Thus, for example, Stadhouders, J. et al., in Neth. Milk Dairy J. 23 (1969) 182 describes the spray-drying of lactic acid bacteria at 70°C, coupled with a post-drying step at 27°C in vacuo. Apparently, preconditioned, i.e. predried, air is not used for the drying. Before the drying, a calcium hydroxide slurry is added to the material to be sprayed. The calcium lactate formed during the spray-drying is advantageous, inasmuch as it is said to have a lower hygroscopicity. In other spray-drying processes known from the prior art, bacterial suspensions to which the most varied types of carrier materials have been added in advance are sprayed. Thus, for example according to SU 724113, a bacterial suspension admixed with dried milk powder, molasses and sodium glutamate is sprayed. According to SU 1616990, a bacterial suspension admixed with the mineral palygorskite is spray-dried. WO-A-88/06181 describes the spray-drying of a bacterial suspension admixed with clay. JP-A-69/67989 describes the spray-drying of yeast cells or bacterial cells which are suspended in a neutral or slightly acidic solution which comprises proteins, carboxymethylcellulose, alginate or alginate ester, disaccharides or higher saccharides or polyhydric alcohols.

The dry microorganism preparations which are known to date from the prior art, in particular those preparations which are used for producing foodstuffs or feedstuffs, have at least one of the following disadvantages:

- 1) the content of viable microbes per unit weight of the dry material is very low owing to the production method, so that large volumes of the dry preparation must be used in the final application;
- 2) the storage stability is too low, so that the dry preparations must be used within a few weeks, if storage under technically complex conditions is impossible;
- 3) the dry preparations have a high dust content, which makes their processing more difficult;

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- 4) the mechanical stability is very low, so that on mixing the preparation with mineral additives, a finely divided abraded material is formed and separation of the solid preparation can be observed;

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- 5) the dissolution rate of the dry preparations is not satisfactory, so that the desired microbiological process for producing the foodstuff or feedstuff only begins slowly and unwanted microorganisms are given the possibility of multiplying, which can lead to considerable losses in quality.

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The production processes known to date from the prior art, in particular the spray-drying processes described to date, are also unsatisfactory for at least one of the following reasons:

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- 1) the processes are technically very complex;
- 20 2) the microorganism survival rate in drying is too low;
- 3) the moisture content of the dry product is too high.

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A first object of the present invention is thus the provision of improved dry microorganism cultures which substantially no longer have the abovementioned deficiencies known from the prior art. In particular, starter cultures which are improved in comparison with the prior art are to be provided. The starter cultures according to the invention are especially to enable improved production of silage.

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A second object of the present invention is the provision of improved processes for producing dry microorganism cultures. In particular, an improved process for spray-drying microorganism cultures should be provided which enables the production of dry preparations having a high content of viable microbes and high storage stability.

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40 The above first object is achieved by providing a dry microorganism culture which comprises at least one microorganism species in carrier-bound form, wherein the culture is present in the form of particles which

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- a) have a particle size of at least about 0.1 mm and
- b) are compressed.

The particulate cultures according to the invention are virtually dust free on account of the chosen particle size. The dust content is preferably in the range from about 0.01 to 0.05% by weight, based on the total weight of the dry culture. This corresponds to a dust index in the range from about 1 to 12 determined gravimetrically by a Casella instrument.

- 10 The particles according to the invention furthermore have a compressed, i.e. compact, structure. This is preferably achieved in their production by a compression step which is explained in more detail below and has not been previously described for dry microorganism preparations. In this operation a preliminary product obtained, for example, by spray-drying, freeze-drying or fluidized-bed-drying (such as the powder concentrate which is obtainable by a spray-drying variant according to the invention and is described below), which usually has a significant dust content (e.g. a dust index from about 25 to 100), is mechanically compressed.

- The compression can be performed, for example, by compacting the pulverulent preliminary product under the action of linear forces, e.g. in the range from about 5 to about 25 kN/cm, in particular from about 10 to about 15 kN/cm, in conventional compacting apparatuses, for example. However, the preliminary product can also be tabletted under the action of pressures in the range from about 50 to about 250 MPa, in particular in the range from about 80 to 200 MPa, such as from about 90 to about 160 MPa, for instance, in conventional tableting presses, for example. Particular preference is given to compression by compacting. In addition, preference is given to compacting powder concentrates obtained according to the invention by spray-drying.

- 35 The provision of microorganism cultures of the type described above surprisingly results in the processing, in particular as starter cultures, being markedly simplified and, moreover, the mechanical stability of the particles and thus the danger of separation of starter culture preparations being markedly decreased. Surprisingly, it has also been found that the compression of the pulverulent preliminary product virtually does not impair product quality with respect to the number of viable microbes. Rather, owing to the high density achieved, the ingress of air and moisture into the dry preparations according to the invention is significantly decreased in such a manner that a considerable improvement in storage stability can be achieved.

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Thus, for example, survival rates of 60% and above after storage for one year at room temperatures were achieved. Advantageous storage stability data of this type have not been described hitherto.

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In particular, the compressed particles can comprise compacted broken material (i.e. material obtained by comminuting with or without classifying compacted product extrudates) having a diameter of from about 0.1 mm to about 2 mm, preferably from 0.3 to 1.25 mm. The diameter here is a value calculated from the total mass distribution of the compressed particles and corresponds to the diameter of spheres of equal mass. The edge length of the particles is in the range from about 0.1 to 2 mm, in particular from about 0.1 to 1.4 mm.

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The compressed particles can, furthermore, be present as tablets of any desired shape, such as round, polygonal or oval, having a diameter of from about 2 to 50 mm and a ratio of diameter to thickness of from about 1:0.1 to about 10:1, in particular from about 1:1 to about 5:1.

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According to a further preferred embodiment of the invention, the dry microorganism cultures comprise, as further component, an effervescence additive, comprising an acid component, such as an organic nonvolatile carboxylic acid, and a gas-forming component, such as a CO₂-forming component. Effervescence formulations of this type have the particular advantage of a surprisingly rapid dissolution after application of the starter culture. As a consequence of this rapid dissolution of the starter culture in its surrounding medium, rapid multiplication of the starter culture microorganisms is ensured, as a result of which losses in quality of the product to be produced using the starter culture can be avoided surprisingly well.

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Preferably, the dry culture compressed according to the invention comprises, as carrier, at least one matrix material for embedding the microorganism cells with or without at least one other additive which stabilizes the cells.

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The carrier used in the dry cultures according to the invention comprises at least one matrix component added as coformulant prior to the drying to usually freshly grown microorganisms, selected from mono-, oligo- and polysaccharides, polyols, polyethers, polymers, such as CMC or PVP, oligo- and polypeptides, from natural sources, such as milk, meat or

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cereals, derived substances or mixed substances, such as sweet whey powder, wheat semolina bran, peptone, alginates, mineral compounds, or mixtures of such matrix substances. In addition, additives having a stabilizing action can be added together with the matrix substance or later, for example antioxidants, such as α -tocopherol or ascorbic acid, or mixtures thereof. Furthermore, a stabilizing action can be exerted by other substances, which are selected from inorganic salts, such as alkali metal chlorides or alkaline earth metal chlorides, inorganic or organic buffers, such as alkali metal phosphate buffer, amino acids, such as aspartic acid or glutamic acid and the salts thereof, organic carboxylic acids, such as citric acid, organic nonvolatile solvents, such as DMSO, and other compounds, such as β -carotene and mixtures of these.

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The microorganism cultures according to the invention preferably comprise viable microorganisms in a concentration of 10^8 to 10^{12} cfu (colony forming units)/g of dry culture. The powder concentrates produced according to the invention comprise from about $5 \cdot 10^8$ to $1 \cdot 10^{12}$, preferably about $4 \cdot 10^{11}$ to $8 \cdot 10^{11}$ cfu/g. The compressed cultures according to the invention comprise from about $1 \cdot 10^{11}$ to $4 \cdot 10^{11}$, in particular about $3 \cdot 10^{11}$ cfu/g. Starter cultures for producing silage comprise from about 1 to $7 \cdot 10^{10}$, in particular about $3 \cdot 10^{10}$ cfu/g.

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In this process the microorganisms can be derived from one microorganism species or a plurality. A particularly preferred species are lactic-acid-producing bacteria, such as those which are suitable for silage production, such as, for example, *Lactobacillus plantarum*.

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For the purposes of the invention, silage comprises feed plant products which have been preserved by the action of microorganisms, for example those based on grass, clover, straw, corn plants, fodder beets, legumes, cereals, such as corn and wheat, and the like.

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The second object of the present invention described above is surprisingly achieved by providing a spray-drying process for producing a dry microorganism culture, comprising at least one microorganism species in carrier-bound form, which comprises

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- a) dissolving or suspending at least one substance suitable for forming a carrier in a liquid comprising at least one microorganism species,
- 5 b) drying the resultant mixture in a spray-dryer, for the spray-drying use being made of a conditioned dried gas heated to a temperature in the range of above about 80°C, in particular from about 90 to about 135°C, preferably from about 100 to about 110°C, such as about 105°C, and
- 10 c) removing the dried material from the spray-dryer, this dried material having an exit temperature of from about 40 to 85°C, in particular from about 45 to 75°C, preferably from about 50 to 65°C, such as about 55°C.
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This spray-drying process according to the invention is also called carrier-bound spray-drying process below. The gas used for the drying is preferably a dried gas having a dew point of below +5°C, in particular having a dew point of from about -10 to about -50°C, such as conditioned compressed air or conditioned nitrogen. For example, compressed air having a dew point of about -25°C and nitrogen having a dew point of about -40°C can be used. A dew point of +5°C is equivalent to roughly 5 g of water per m³ of air.

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25 According to a preferred embodiment of the spray-drying process according to the invention, in a downstream further stage d), the dried material is subjected to a post-drying. The post-drying temperature is in the range of from about 15 to 50°C, such as from about 25 to 40°C. The post-drying is performed, for example, in a gas atmosphere or in vacuo; alternatively to this, there is also the possibility of mixing a desiccant homogeneously with the dry microorganism preparation obtained in accordance with stage c).

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35 Because of its design, the spray-drying process according to the invention surprisingly permits microorganism suspensions to be dried at survival rates of up to 100%. Owing to the use of conditioned gas in the spray-drying as well as the optional post-drying step, surprisingly, dry preparations having an extremely low moisture content (of from about 2 to 3% by weight of water), corresponding to a water activity a_w of from 0.03 to 0.15, are provided. This directly causes the microorganism cultures which have been spray-dried according to the invention, with or without post-drying, to have survival rates of up to 60% after storage for 1 year at ambient temperature and ambient air conditions.

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Owing to the surprisingly high survival rate in the above-described spray-drying, the content of viable microorganisms is markedly high. The resultant spray-dried product is therefore also called powder concentrate and, to
5 reduce the concentration of viable cells, can be further diluted, depending on the field of application. The powder concentrate is particularly suitable for preparing the above-described compressed particulate cultures according to the invention.

10 The present invention therefore also relates to a process for producing the above-described compressed microorganism cultures, which comprises

15 i) producing a powder concentrate of the microorganism culture by carrier-bound spray-drying, carrier-bound freeze-drying or carrier-bound fluidized-bed drying,

20 ii) with or without admixing the powder concentrate with one or more coformulants and

iii) compressing this mixture by compacting or tableting.

25 Preferably, in a further process step, the compressed mixture is broken, i.e. comminuted, and may be classified to give compressed granules of the desired size using a screen of suitable mesh width.

30 The present invention further relates to a process for producing a dry agglomerated microorganism culture, which comprises

35 i) producing a powder concentrate of the microorganism culture by carrier-bound spray-drying, carrier-bound freeze-drying or carrier-bound fluidized-bed drying,

ii) with or without admixing the powder concentrate with one or more coformulants and

40 iii) compressing this mixture by agglomeration.

Carrier-bound means here the presence of at least one matrix material of the above-described type during drying.

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According to a preferred embodiment of the above-described compacting process or tableting process or agglomeration process, stage i) is carried out in particular in accordance with the above-described spray-drying process.

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The product obtained by the above-described compression processes is, for the purposes of the present invention, also called compressed or compacted dry concentrate (in the cfu range from about $1 \cdot 10^{10}$ to $1 \cdot 10^{11}$) and can be marketed as such, e.g. as a concentrated starter culture.

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The present invention further relates to the use of the compressed dry microorganism cultures according to the invention as starter cultures for producing foodstuffs, such as for the production of milk products, such as sour cream, buttermilk, yogurt, kefir, coumis, curd cheese, for producing sourdough, uncooked sausage, and for producing feedstuffs, such as silage. For this purpose, the culture, with or without dissolution, is mixed with the foodstuff substrate or feedstuff substrate. If the cell count in the starter culture should be too high, it may be diluted, e.g. by mixing with an inert solid, such as lime, in particular feed lime.

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The present invention further relates to foodstuffs and feedstuffs which have been produced using the starter cultures according to the invention.

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The present invention is described in more detail in the sections now following with reference to the accompanying figure.

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Figure 1 shows diagrammatically a possible way of producing, from powder concentrate, granules compacted in accordance with the invention.

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Usable microorganisms

The present invention is not restricted in principle to certain microorganism cultures. Rather, those skilled in the art recognize that the present invention is applicable to any microorganisms, in particular bacteria and yeasts, which can be converted to a dry microorganism preparation under the conditions specified in the present description. A suitable group of microorganisms which can be used according to the invention are the group of lactic-acid-producing bacteria. In particular, these are bacteria which are suitable for the homofermentative lactic

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acid fermentation, i.e. break down glucose to lactate via the fructose bisphosphate pathway. Typical representatives of this group are bacteria of the genera *Lactobacillus* sp., *Streptococcus* sp. and *Pediococcus* sp. Concrete examples of lactobacilli which
5 may be mentioned are *Lactobacillus bulgaricus*, *Lactobacillus acidophilus*, *Lactobacillus helveticus*, *Lactobacillus bifidus*, *Lactobacillus casei*, *Lactobacillus lactis*, *Lactobacillus delbrueckii*, *Lactobacillus thermophilus*, *Lactobacillus fermentum*, *Lactobacillus brevis* and *Lactobacillus plantarum*. Examples of
10 suitable streptococci are *Streptococcus lactis*, *Streptococcus cremoris*, *Streptococcus diacetylactis*, *Streptococcus thermophilus*, *Streptococcus pyrogenes*, *Streptococcus salivarius*, *Streptococcus faecalis*, *Streptococcus faecium*; and examples of suitable pediococci are *Pediococcus cerevisiae* and *Pediococcus*
15 *acidilactici*.

Fermentation of the microorganisms

20 To carry out the present invention, preferably, use is made of freshly prepared microorganism suspensions. The fermentation media or fermentation conditions optimum for each microorganism are either known from the prior art or can be determined in only a few routine experiments by the person skilled in the art who is
25 entrusted with the culture of microorganisms.

However, usually, the fermentation is carried out in such a manner that starting from a liquid or semi-solid preliminary
30 culture (culture volume from about 10 to 200 ml), freshly prepared sterile fermentation medium is inoculated under sterile conditions, where the volumetric ratio of preliminary culture to culture medium can be from about 1:50 to 1:200. Preferably, freshly grown preliminary cultures are used which are in a late
35 phase of logarithmic growth. Depending on the microorganism being grown, they are cultured under specific optimized growth conditions (such as temperature and pH). Usually, the growth temperature is in the range from about 20 to 40°C, but, for example when thermophilic bacteria are being grown, markedly
40 higher temperatures can be present. The fermentation batch is kept uniformly agitated, for example by moderate stirring or introducing air or nitrogen in order to prevent the development of temperature or substance gradients and to ensure continuous growth in this manner. After the growth phase is complete
45 (determined for example by achieving a defined cell density or consumption of one of the added nutrients), the cell suspension

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can be used directly to produce the dry preparations according to the invention.

- 5 However, it is also possible to concentrate the resultant original cell suspension to increase the cell count. Suitable methods for this are, for example, centrifugation, ultrafiltration or thin-film evaporation. However, a centrifugation step is usually used to concentrate the cell
- 10 suspension, which centrifugation step is preferably carried out at a decreased temperature, that is to say in the range from about 4 to 10°C.
- 15 Instead of the concentration, or in combination with it, there is also the possibility of subjecting the freshly cultured cell suspension to a washing step in order to remove culture constituents, such as metabolic products, which may have an adverse effect on the activity. In this case, the procedure
- 20 usually adopted is that, preferably at from about 4 to 10°C, the original culture broth is first concentrated to give a suspension of high cell density and this is then taken up in a suitable buffer solution and diluted to the desired cell density. If necessary, the washing step can also be repeated a plurality of
- 25 times. Solids contents which can be used according to the invention of cultures of microorganisms suitable for producing the dry preparations according to the invention are usually in the range from about 5 to 25% by weight, such as from about 10 to 20% by weight.
- 30
- The microorganisms can be cultured by batch fermentation or continuously.
- 35 To further illustrate the invention, in the section below, a more detailed description is given of culturing a lactic acid bacterium, in particular *Lactobacillus plantarum*. This is a bacterium which is to be found in particular on intact and decomposing plants and is particularly suitable for producing
- 40 silage feedstuffs.

- A suitable fermentation medium comprises, per liter of medium, from about 40 to 60 g of glucose, from about 30 to 60 g of yeast extract and a cocktail of customary trace elements, such as
- 45 magnesium, manganese and, optionally, iron. The pH of the fermentation medium is from about 6 to 7. The fermentation temperature is from about 33 to 38°C. The pH of the fermentation

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medium can be kept within the desired range by adding sterile sodium hydroxide solution. Growth is complete when glucose consumption or lactic acid synthesis can no longer be observed.

- 5 According to a lactobacillus fermentation variant which is suitable according to the invention, after about 80% or 90% of the growth is achieved, the fermentation medium temperature is increased to from about 42 to 46°C until the added glucose is completely consumed. It has been found according to the invention
10 that cultures produced in this manner are particularly stable in particular in the spray-drying, as a result of which high survival rates are achievable. Comparable growth variants are also conceivable with other microorganisms which can be used according to the invention.
15

- After growth is complete, the fermentation batch is brought to the desired cell density. If desired, the cell suspension can be washed until it is virtually lactate free. The cell count of a
20 microorganism suspension suitable according to the invention is usually in the range from about 1×10^{10} to about 5×10^{12} cfu/g of suspension.

Carrier substances

- 25 The dry microorganism cultures prepared according to the invention, in addition to any nonvolatile constituents present from the respective fermentation batch, such as metabolic
30 products, comprise at least one matrix material with or without other stabilizing substances. These coformulants are preferably selected from inorganic salts or buffers, at least one other compound which is selected from mono-, oligo- and
polysaccharides, polyols, polyethers, amino acids, oligo- and
35 polypeptides, milk-derived compounds, organic carboxylic acids, mineral compounds, organic carrier materials such as wheat semolina bran, alginates, DMSO, PVP (polyvinylpyrrolidone), CMC (carboxymethylcellulose), α -tocopherol, β -carotene and mixtures thereof.

- 40 Examples of suitable saccharide carrier components are sucrose, fructose, maltose, dextrose, lactose and maltodextrin. An example of a suitable polyol is glycerol. Examples of suitable amino
45 acids are glutamic acid, aspartic acid and the salts thereof. An example of a suitable peptide carrier is peptone. An example of a milk-derived compound is, in addition to the abovementioned

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maltodextrin, also sweet whey powder. Suitable organic carboxylic acids are, for example, citric acid, malic acid and L-ascorbic acid. Examples of suitable mineral carriers are montmorillonite and palygorskite.

5

However, preferably, as carrier for the dry microorganism preparations according to the invention, use is made of mixtures of the abovementioned classes of substances. Mixtures of this type preferably comprise, as main component, a matrix material, such as one of the abovementioned saccharide components or, for example, sweet whey powder, with or without a minor content of at least one further component, such as a buffer component (for example citric acid) or an antioxidant (for example L-ascorbic acid or α -tocopherol). The addition of further stabilizing constituents, such as sodium glutamate and/or peptone, has likewise proved to be advantageous.

The matrix component is customarily used in carrier compositions usable according to the invention in about 5 to 30 times the amount of the other carrier constituents. Examples of particularly suitable carrier combinations are:

a) sweet whey powder/citric acid/L-ascorbic acid (weight ratio about 40:1:1).

b) maltodextrin/lactose/citric acid/L-ascorbic acid (weight ratio about 20:20:1:1), unsupplemented or supplemented by about 1.5 parts of β -carotene and 0.5 part of α -tocopherol per part of citric acid.

c) maltodextrin/sodium glutamate/L-ascorbic acid (weight ratio about 10:1.5:1).

d) lactose/glucose/peptone/citric acid (weight ratio about 6:6:1.2:1).

40

The carrier substances according to the invention can be added to the microorganism suspension either as solid or in dissolved form. However, preferably, a sterile solution of the carrier/carriers is prepared, this is cooled to a temperature of from 4 to 10°C and this is mixed with the likewise cooled microorganism suspension with gentle stirring. To prepare a homogeneous suspension, the resultant mixture is stirred with

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further cooling for a period of from about 10 minutes to 1 hour.

Preparation of dry microorganism preparations

5

The microorganism suspension containing the carrier added in the manner described above can then be dried in various ways.

Suitable drying processes are in principle freeze drying, fluidized-bed drying and, preferably, spray-drying. For the

10 purposes of the present invention, spray-drying also comprises modified spray-drying processes, such as spray-agglomeration or agglomerating spray-drying. The latter process is also known under the name FSD (fluidized spray-dryer) process.

15 Freeze-drying for preparing dry microorganism cultures according to the invention can be carried out, for example, on the basis of the freeze-drying process described in EP-A-0 259 739 or US-A-3 897 307. The contents of these publications are hereby incorporated completely by reference.

20

A suitable fluidized-bed drying process is described, for example, in the dissertation by U. Kessler on the subject "Experimentelle Untersuchung und Modellierung der Überlebensrate von Milchsäurebakterien bei der thermischen Trocknung"

25 [Experimental study and modeling of the survival rate of lactic acid bacteria during thermal drying], Technical University of Munich, 1993. The contents of this publication are likewise incorporated completely by reference. To carry out the

30 fluidized-bed drying process, it is advantageous that the carrier material to be used, in particular the matrix component, is introduced in a fluidized bed and this is sprayed with the microorganism suspension in the manner described by U. Kessler.

35 However, the drying process which is most preferred according to the invention is spray-drying. Those methods which can be used according to the invention are essentially all spray-drying techniques known hitherto. The material to be sprayed can, for example, be dried cocurrently or countercurrently; spraying can
40 be carried out by means of a single-component or multiple-component nozzle or by means of an atomizer wheel.

Preference is given according to the invention to the use of
45 material to be sprayed having a solids content (after addition of the carrier) of from about 10 to 40, such as from about 10 to 25%

by weight.

The spray-drying process according to the invention is carried out in such a manner that a conditioned dry gas having a temperature in the range of above about 80°C is introduced into the drying apparatus. In particular, the inlet temperature should be in the range of from about 90 to 135°C. Particular preference is given to a drying temperature in the range of about 105°C. The rate of the drying process is designed according to the invention in such a manner that the exit temperature of the drying material from the dryer is in the range of about 45 to 75°C, in particular in the range of from about 50 to 65°C, preferably about 55°C.

Of particular importance to the process according to the invention is the use of preconditioned, i.e. low-moisture, drying air. Preferably, use is made of compressed air having a dew point at about -25°C.

The drying process according to the invention shall be carried out in such a manner that a very low residual moisture content is present in the dry material. Preferably, the water activity a_w in the drying material should be less than 0.4. However, to further improve the long-term storage stability, according to the invention water activities of less than 0.15, preferably in the range from about 0.03 to 0.1 are sought after. The percentage water content is preferably from about 2 to 3% by weight. Most preferably, this is achieved by adding a post-drying step subsequently to the spray-drying step. The drying material for this purpose is, for example, post-dried in a fluidized bed, preferably at a temperature in the range of from 15 to 50°C, for a period of, for example, from 15 minutes to 20 hours. Again, preferably, conditioned compressed air or conditioned nitrogen serves as drying gas. However, the post-drying can also be performed by applying a vacuum of from about 1 to 50 mm Hg for a period of from about 15 minutes to 20 hours and at a temperature of from about 15 to 50°C. In this case, preference is given to stirring the drying material, for example, using a paddle agitator.

Instead of the above-described physical post-drying processes, it is also conceivable to add specific desiccants to the dry material obtained from the spray-drying. Desiccants of this type should themselves have a very low water activity, such as an a_w of 0.01 or less. Examples of suitable desiccants are inorganic salts, such as calcium chloride and sodium carbonate, organic

polymers, such as the product obtainable under the trade name Kollidion 90 F, and silicon-dioxide-containing desiccants, such as silica gel, zeolites and desiccants which are obtainable under the trade name Tixosil 38, Sipernat 22 S or Aerosil 200.

5

According to the invention, it was surprisingly found that, despite the relatively high drying temperatures, the survival rate for the dry preparations according to the invention had excellent values, namely of $75\% \pm 25\%$.

10

The content of viable microorganisms is in the range of from about 5×10^8 to 1×10^{12} cfu/g of dry matter. These preparations are also called according to the invention powder concentrates.

15 Since, for individual final applications, lower contents of viable microorganisms are also completely sufficient, powder concentrates of this type can therefore if appropriate be blended to the final count of viable microorganisms by mixing with further inert carrier material.

20

Preparation of compressed dry microorganism cultures

The powder concentrates obtainable by the above-described drying
25 processes usually have a relatively high dust content and are thus not yet satisfactorily handleable for individual applications. Furthermore, various applications require an increased mechanical stability of the dry cultures. It is therefore necessary to improve the properties of the
30 above-described powder concentrates by a further compression step.

To reduce the dust content of the powder concentrates according to the invention, it is possible to agglomerate these in a
35 conventional manner to form granules, or using external forces, to compact them or tablet them.

Agglomeration is a generally known process, and is described, for
40 example, by Schade, A. and Leuenberger, H. in Continuous fluidized-bed spray granulation, Chemie Ingenieur Technik (1992) 64 (1992) 1016; Uhlemann, H., Preparation of pharmaceutical granules in a combined wet granulation and multichamber fluidized-bed drying process, Chemie Ingenieur Technik 62 (1990),
45 822; or Rosch, M. and Probst R., Granulation in the fluidized bed, Verfahrenstechnik (1975), 9, 59.

Use can be made according to the invention of agglomeration using a mixer. For this purpose, the above-described powder concentrate is charged into the mixer and oil, water or an aqueous or alcoholic solution of sugars, polymers or other additives is
5 sprayed in to agglomerate the powder concentrate.

In addition, use can be made according to the invention of agglomeration in a fluidized bed. In this case, powder
10 concentrate is vortexed with gas feed and sprayed with an aqueous or alcoholic solution of sugars, polymers or other additives to form the agglomerate. Suitable processes for this purpose are described, for example, in WO-A-88/06181, in the dissertation by U. Kessler (loc. cit.) and by K. Fuchs in ZFL 45 (1994) 31. The
15 disclosure of the abovementioned publications is hereby incorporated by reference.

Agglomeration produces granulated microorganism cultures having a particle size in the range of from about 0.1 to about 4 mm, in
20 particular from about 0.3 to 2.5 mm.

However, particularly preferred according to the invention is the preparation of dry microorganism cultures which are present in the form of particularly highly compressed particles. This is
25 carried out according to the invention either by tableting in conventional tablet presses or with the use of conventional compacting apparatuses equipped with two counter-rotating rolls.

30 To compress the powder concentrates obtainable according to the invention, to these are usually added one or more coformulants or additives to modify the processability to the end product or the properties of the end product.

35 To improve the flowability of the powder concentrate, a free-flowing agent is preferably added. Examples of a suitable free-flowing agent are spray-dried silicon dioxide powders, which are obtainable, for example, under the trade name Sipernat 50. To improve the storage stability of the solid formulations according
40 to the invention, in addition, conventional antioxidants, such as L-ascorbic acid, can be added. Furthermore, desiccants of the above-described type can additionally be added.

45 The action of the cultures according to the invention is markedly improved if measures are taken which, after the culture has been applied, lead to a rapid breakdown of the grain structure and

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thus to a rapid release of the microorganisms. One possibility of achieving this is the addition of a readily water-soluble component which thus accelerates the breakdown of the grain structure. Suitable compounds are, for example, poly(ethylene glycol)s, which are obtainable, for example, under the trade name Pluriol E.

Another solid formulation particularly preferred according to the invention comprises what is termed an effervescence additive. This is a gas-releasing component, in particular a CO₂ source, for example an alkaline earth metal hydrogen carbonate, preferably sodium hydrogen carbonate or ammonium hydrogen carbonate; and an acid component, preferably selected from citric acid, ascorbic acid or malic acid. This effervescence additive, in the presence of moisture, produces a spontaneous gas formation with breakdown of the grain structure and rapid release of the microbial cells.

In particular, to prepare highly compressed, compacted or tabletted microorganism cultures, it is advisable to add compacting or tableting aids. This is because it has surprisingly been found according to the invention that adding such compacting aids decreases the pressures acting on the microorganisms during the compacting and thus markedly improves the survival rate of the microbes. Examples of suitable compacting aids are microcrystalline cellulose, sugars and mixtures thereof. Concrete examples of microcrystalline cellulose are products which are commercially available under the trade names Avicel, Arbocel and Vivapur. Examples of suitable sugars are maltose, maltodextrin and lactose preparations, which are obtainable under the trade names Granulac, Tablettose or FloLac. An example of a suitable mixed cellulose/sugar product is the commercial preparation Cellactose. A further suitable tableting aid is a lactose preparation granulated using PVP, obtainable under the trade name Ludipreß.

Other suitable additives are poly(ethylene glycol)s (Mw from 100 to 10,000) which can have a stabilizing action on the cells embedded in the matrix.

The accompanying Figure 1 shows a flow diagram for the further processing according to the invention of the powder concentrates to give a compacted product according to the invention. Powder concentrate PK is mixed in the mixer M1 with the coformulants or additives ZU, passes from there into a reservoir B1 which feeds

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the compactor A1. The product ribbon exiting from the compactor is precomminuted or finely comminuted in the grinders Z1 and Z2 and in the screen F1, product PR is separated off from dust fractions having a particle size of less than 0.3 mm. This

- 5 material is fed to the mixer M1 as recycled material RÜ. The product PR having a particle size of 0.3 mm or above, such as from 0.3 to 1.5 mm, passes to the packaging station or may be subjected to further processing, such as a coating process.

10

Suitable coating materials, which preferably are additionally to hinder the ingress of moisture to the dry preparation, are, for example, alcoholic solutions of PVP, in particular a PVP product which is commercially available under the trade name Kollidon

- 15 VA64. Another usable coating system is a mixture of shellac and Kollidon 25 or 30, which is supplemented with titanium dioxide and tallow and is likewise present in alcoholic solution.

- 20 To reduce the cell count further if necessary, a coated or uncoated product obtained in this manner can be blended, for example, with lime, or another suitable mineral additive.

Examples

25

Analytical methods used in the following examples:

a) Cell count determination:

30

Cell counts are determined in the conventional manner by serial dilution with sterile 0.9% strength NaCl solution and subsequent plating on MRS agar (Difco Laboratories). Colony-forming units (cfu) were counted after incubation for 48 hours at 37°C. Only

35 plates which contained between 30 and at most 300 colonies were evaluated. Generally, 3 plates per stage were evaluated and the mean taken.

- 40 The specific cell count of a sample was determined by calculation, dividing the cell count per gram of sample by the relative sample dry matter content.

b) Determination of the survival rate on drying:

45

20

The survival rate during drying was calculated from the specific cell count of the sample before drying divided by the specific cell count after drying. It was always expressed in percent.

5 c) Determination of storage stability:

To determine the storage stability of a dried sample, the specific cell count of the dried sample was determined immediately after drying (day_0). The dried cell material was stored under air in an opaque tightly sealed vessel at room temperature ($21^\circ\text{C} \pm 2^\circ\text{C}$) for extended periods. The specific cell count was determined again at regular intervals (day_N). The storage stability was calculated from the quotient of specific cell count day_N /specific cell count day_0 .

If the specific cell count after drying was, for example, $5 \cdot 10^{11}$ cfu/g of DM and, after storage for 8 weeks, $4 \cdot 10^{11}$ cfu/g of DM, the storage stability was 80% of the initial value.

20

d) Moisture content measurement:

Electronic moisture analyzer HR 73 from Mettler

Procedure: approximately 2 g of powder are distributed onto the measuring scales of the instrument. Measurements are taken at a drying temperature of 105°C up to constant weight (switch-off criterion: max. 1 mg of weight loss in 50 seconds).

30 e) Measurement of water activity:

Hygroscope DT instrument from Rotronic AG, Zürich, Switzerland

The product is placed in the sample holder and this is positioned in the measuring chamber thermostatted to 25°C . After closing the measuring chamber and an equilibration time of 20 minutes, the instrument measurement value is read off.

f) DSC measurement to determine the glass transition temperature T_g :

40

TA4000 instrument from Mettler

Sample weight approximately 15 mg, heating rate $20^\circ\text{C}/\text{min}$, samples were flushed with a nitrogen stream during measurement.

45 DSC = Differential Scanning Calorimetry

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Microorganism culture examples

Example K1: Batch Fermentation 10 liter scale

- 5 10 l of a fermentation medium which comprised the following constituents were placed in a 14 l fermenter and sterilized at 121°C for 30 minutes:

10	Glucose monohydrate	550.0 g
	50% yeast extract suspension (pH 4.5 with phosphoric acid)	750.0 g
	Tween 80®	10.0 g
15	MgSO ₄ * 7 H ₂ O	5.0 g
	MnSO ₄ * 1 H ₂ O	0.5 g

- After sterilization, the medium was adjusted to pH 5.8 at 37°C using sterile 25% strength sodium hydroxide solution and the
20 medium was blanketed with a gentle stream of sterile nitrogen. The fermenter was stirred at 150 rpm.

- The fermenter was then inoculated with 100 ml of a preculture of Lactobacillus plantarum (BASF strain LU 3244) which had
25 previously been grown for 16 h at 37°C in MRS nutrient medium (Difco Laboratories). The culture pH was continuously kept at 6.2 using 25% strength sodium hydroxide solution.

- 30 The course of the fermentation was followed from the sodium hydroxide solution consumption. As soon as no more sodium hydroxide solution was consumed (total consumption 890 g), all of the fermentation broth was drained off and centrifuged at 8°C. The biomass was resuspended in about 600 g of supernatant and made up
35 to exactly 1000 g with supernatant. The dry matter content was determined using an infrared drying balance (105°C to constant weight). The solids content of this suspension was 15%.

Example K2: batch fermentation 200 liter scale

- 40 180 l of a fermentation medium which comprised the following constituents were placed in a 200 l fermenter and sterilized at 121°C for 30 minutes:

45	Glucose monohydrate	11 kg
	50 % yeast extract suspension	15 kg

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Tween 80®	200 g
MgSO ₄ * 7 H ₂ O	200 g
MnSO ₄ * 1 H ₂ O	10 g

5

After sterilization, the medium was adjusted to pH 5.8 at 37°C using sterile 25% strength sodium hydroxide solution and the medium was blanketed with a gentle stream of sterile nitrogen.

- 10 The fermenter was then inoculated with 2000 ml of a preculture of *Lactobacillus plantarum* (3244) which had previously been grown for 24 h at 30°C in MRS nutrient medium. The pH of the culture was continuously controlled using 25% strength sodium hydroxide solution.

15

The course of the fermentation was followed from the sodium hydroxide solution consumption. In total, 16.43 kg of 25% strength NaOH were consumed. As soon as sodium hydroxide solution was no longer consumed, all of the fermentation broth was drained off and harvested at 8°C using a continuous separator. The harvested biomass had a weight after centrifugation of 20 kg, and the solids content of this suspension was 12.3%. The cell count of the suspension was $1.04 \cdot 10^{11}$ cfu/g of suspension. The specific cell count was $8.45 \cdot 10^{11}$ cfu/g of dry matter (DM).

25

Example K3: batch fermentation with temperature shock

A fermentation was carried out in a similar manner to Example 2.

- 30 At a sodium hydroxide consumption corresponding to 90% of the expected value, the fermenter temperature was increased to 44°C and kept until all of the sugar present had been consumed. The cells were then harvested as described in Example K2. The cell count of the fermentation broth was $1.8 \cdot 10^{11}$ cfu/g at a solids content of 21.17%. This corresponds to a specific cell count of $8.5 \cdot 10^{11}$ cfu/g DM.

35

Example K4: continuous fermentation

- 40 10 l of a fermentation medium having the following composition were charged into a 14 l fermenter and sterilized at 121°C for 30 minutes (production fermenter):

- | | |
|---|---------|
| 45 Glucose monohydrate | 400.0 g |
| 50% yeast extract suspension
(pH 4.5 with phosphoric acid) | 500.0 g |

23

	KH ₂ PO ₄	30.0 g
	Citric acid monohydrate	21.0 g
	Tween 80®	10.0 g
5	MgSO ₄ *7 H ₂ O	5.0 g
	MnSO ₄ *1 H ₂ O	1.7 g
	(NH ₄) ₂ Fe(SO ₄) ₂ *6 H ₂ O	0.4 g

10

2000 l of the same medium were charged into a second fermenter having a total volume of 3000 l and sterilized (reservoir fermenter). Both fermenters were connected by a sterilizable line. Via an intermediate vessel which stood on a balance, using

15

an automatic control system, 3 l of fresh medium were pumped every hour into the production fermenter. The temperature of the production fermenter was controlled to 37°C. The pH was controlled to 5.8 using 25% strength NaOH. The fermenter was stirred at 150 rpm and blanketed with nitrogen at 0.1 VVM.

20

Via a second pump, 3 l of medium were continuously taken off every hour and collected in a stainless steel collection vessel precooled to from 0 to 4°C. The biomass concentration was determined by turbidimetry and was 3.5 g/l. The glucose

25

concentration in the production fermenter effluent was, after the initial growth phase, 0 g/l at all times. The cell count of the fermentation broth was $1.48 \cdot 10^{10}$ cfu/g of broth. The dry matter content of the fermentation broth was 6.89 %, equivalent to 217 g DM. The specific cell count was $2.15 \cdot 10^{11}$ cfu/g of DM.

30

Example K5: cell harvest with washing step to remove sodium lactate

35

72 l of fermenter discharge from Example K4 were harvested continuously at 8°C using a commercial separator. About 7 kg of cell suspension was obtained. To this was added a washing solution which comprised 40 l of deionized water, 450 g of NaCl and 136 g of KH₂PO₄. The pH of the washing solution had been

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adjusted in advance to 7.0 using 25% strength sodium hydroxide solution. The about 50 l of resuspended cells were again separated. 3160 g of concentrated washed cell suspension were obtained. The solids content of the suspension was 9.97%. The cell count was $2.49 \cdot 10^{11}$ cfu/g of suspension. The specific cell

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count was $2.5 \cdot 10^{12}$ cfu/g DM.

24

This washed cell suspension was virtually free of sodium lactate. The biomass concentration was determined by turbidimetry to be 80 g/l.

5 **Examples of preparation by spray-drying of powder concentrates according to the invention**

10 The spray-drying experiments described in the following section for preparing powder concentrates according to the invention are carried out in a laboratory spray-dryer of type Niro Minor from Niro, Copenhagen, Denmark. The ready-to-spray bacterial suspension is sprayed via a two-component nozzle cocurrently with preconditioned heated compressed air into the plant drying tower, 15 the dried product is separated from the air using a cyclone and collected.

Example S1

20 To prepare a coformulant solution, 200 ml of deionized water (completely demineralized water) are heated to 60°C. 150 g of sweet whey powder, 7.5 g of NaCl, 3.8 g of KH_2PO_4 , 3.8 g of citric acid and 3.8 g of L-ascorbic acid are dissolved therein, adjusted to pH 7 using 40% strength aqueous NaOH and made up to 400 g 25 total mass using deionized water. This solution is cooled to 5°C.

200 ml of washed, i.e. essentially sodium-lactate-free centrifuged ferment (prepared in a similar manner to Example K5) (12.7% solids content (S.C.)) are placed in an ice bath at a 30 temperature of 5°C and 400 g of coformulant solution, cooled to 5°C, are added with stirring. The mixture of centrifuged ferment and coformulants is further stirred for 30 minutes at 500 rpm using a magnetic stirrer with ice bath cooling. By means of spray-drying (Niro Minor apparatus) the mixture is then converted 35 into a powder concentrate A, which is separated off in the cyclone. In the course of this, the reservoir from which the mixture is metered is cooled to 4°C, the inlet temperature is from 105 to 110°C, the exit temperature is from 53.5 to 55.5°C. A two-component nozzle is used, conditioned air (dew point -25°C) at 40 4 bar being used to spray the mixture of centrifuged ferment and coformulants.

The powder concentrate A is further dried at room temperature for 45 2 hours in a nitrogen-operated (dew point = -40°C) fluidized bed, powder concentrate B being obtained.

25

Characterizations:

Ready-to-spray mixture: 35% S.C., $2.84 \cdot 10^{11}$ cfu/g of dry matter

Powder concentrate A: water activity $a_w = 0.135$

- 5 Powder concentrate B: water activity $a_w = 0.076$,
moisture content 3.4%,
 T_g from DSC measurement: 54°C ,
 $1.98 \cdot 10^{11}$ cfu/g of dry matter (equivalent
10 to 70% survival rate in the drying)

Storage study of powder

concentrate B: cfu counts with room-temperature storage
in containers sealed under ambient

- 15 air: $2.0 \cdot 10^{11}$ cfu/g of dry matter (100%)
after 30 days

Example S2

- 20 To prepare a coformulant solution, 200 ml of deionized water are
heated to 70°C . 75 g of maltodextrin (Glucidex IT6, Roquette),
75 g of lactose, 7.5 g of NaCl, 3.8 g of KH_2PO_4 , 3.8 g of citric
acid and 3.8 g of L-ascorbic acid are dissolved therein, the
25 mixture is adjusted to pH 7 using 40% strength aqueous NaOH and
made up to 400 g total mass using deionized water. This solution
is cooled to 5°C .

- 200 ml of washed, i.e. essentially sodium-lactate-free
30 centrifuged ferment (16.5% S.C.; prepared similarly to example
K5) are mixed, at 5°C , with stirring into 400 g of coformulant
solution, cooled to 5°C . The mixture is stirred for 30 minutes at
250 rpm by a magnetic stirrer with ice bath cooling. 101 ml of a
solubilized mixture prepared in accordance with EP-A-0 479 066
35 (BASF) from 25% Tween 80, 5% β -carotene and 2% α -tocopherol are
then added and further stirred for 10 minutes with ice bath
cooling. This mixture is then converted by spray-drying, as
described in Example S1, into a powder concentrate A (inlet
temperature 105°C , exit temperature from 54 to 55°C). The powder
40 concentrate A is not further dried.

Characterizations:

Ready-to-spray mixture: 29% S.C., $3.84 \cdot 10^{11}$ cfu/g of dry matter

- 45 Powder concentrate A: water activity $a_w = 0.065$,
moisture content 2.8 %, T_g from DSC measurement: 61°C ,

26

2.22·10¹¹ cfu/g of dry matter (equivalent
to 58% survival rate in the drying)

Storage study on powder

- 5 concentrate A: cfu counts for room-temperature storage in
containers sealed under ambient air:
1.9·10¹¹ cfu/g of dry matter (86%)
after 30 days

10

Example S3

- 400 ml of unwashed, i.e. sodium-lactate-containing, centrifuged
ferment (prepared similarly to Example K4)(14.3 % S.C.) are
15 placed in an ice bath at a temperature of 5°C. 57.2 g of Glucidex
IT6, 8.6 g of L-ascorbic acid and 5.7% of sodium glutamate are
stirred as solids into the cooled centrifuged ferment with
stirring at 700 rpm by means of a magnetic stirrer. The pH is
adjusted to 7 using 40% strength aqueous NaOH. The mixture of
20 centrifuged ferment and coformulants is further stirred for 30
minutes at 500 rpm using a magnetic stirrer at approximately 3°C
with ice bath cooling. The mixture is then converted by
spray-drying, as described in Example S1, into a powder
concentrate A (inlet temperature 105°C; exit temperature from 54.5
25 to 55.5°C).

- The powder concentrate A is further dried at room temperature in
a nitrogen-operated fluidized bed for 2 hours, a powder
concentrate B being obtained.

30

Characterizations:

- Ready-to-spray mixture: 27% S.C., 4.65·10¹¹ cfu/g of dry matter
Powder concentrate A: water activity a_w = 0.197
35 Powder concentrate B: water activity a_w = 0.072,
moisture content 3.8%,
 T_g from DSC measurement: 52°C,
4.64·10¹¹ cfu/g of dry matter (equivalent
40 to 100 % survival rate in the drying)

Storage study on powder concentrate B:

- cfu counts with room-temperature storage
in containers sealed under ambient air:
45 4.1·10¹¹ cfu/g of dry matter (88%)
after 28 days

Example S4

215 ml of washed, i.e. essentially sodium-lactate-free,
5 centrifuged ferment (prepared similarly to Example K5) (14.5%
S.C.) are placed in an ice bath at a temperature of 5°C. 31.2 g of
Glucidex IT6, 4.7 g of ascorbic acid and 3.1% of sodium glutamate
are then stirred in as solids into the cooled centrifuged ferment
with stirring at 700 rpm by a magnetic stirrer. The pH is
10 adjusted to 7 using 40% strength aqueous NaOH. The mixture of
centrifuged ferment and coformulants is further stirred for 30
minutes at 500 rpm using a magnetic stirrer with ice bath
cooling. The mixture is then converted by spray-drying, as
described in Example S1, into a powder concentrate A (inlet
15 temperature 105°C; exit temperature from 54.5 to 55.5°C).

The powder concentrate is further dried at room temperature in a
nitrogen-operated fluidized bed for 2 hours, powder concentrate B
20 being obtained.

Characterizations:

Ready-to-spray mixture: 28% S.C., $8.76 \cdot 10^{11}$ cfu/g of dry matter

25 Powder concentrate B: water activity $a_w = 0.044$,
moisture content 3.8%,
 T_g from DSC measurement: 48°C,
 $7.17 \cdot 10^{11}$ cfu/g of dry matter (equivalent
to 82% survival rate in the drying)

30 Storage study on powder

concentrate B: cfu counts for room-temperature storage in
containers sealed under ambient air:
3.7 · 10¹¹ cfu/g of dry matter (52%)
35 after 27 days

Example S5

40 To prepare a coformulant solution 1, 40 ml of deionized water are
charged and 33.3 g of lactose and 6.3 g of peptone are dissolved
therein, the mixture is made up to a total mass of 83 g with
deionized water and adjusted to pH 7 using 40% strength aqueous
NaOH. To prepare a coformulant solution 2, 40 ml of deionized
45 water are charged and 33.3 g of glucose-1-hydrate and 5.4 g of
citric acid are dissolved therein, the mixture is made up to a
total mass of 83 g with deionized water and adjusted to pH 7

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using 40% strength aqueous NaOH. These solutions 1 and 2 are cooled to 5°C.

200 ml of washed, i.e. essentially sodium-lactate-free,
5 centrifuged ferment (prepared similarly to Example K5) (12.7% S.C.) are mixed with 83 g of the cooled coformulant solution 1 in an ice bath at approximately 4°C. The mixture is stirred for 30 minutes with ice bath cooling. 83 g of the cooled coformulant solution 2 are then added with stirring and further stirred for
10 30 minutes with ice bath cooling. Then this mixture is converted by spray-drying, as described in Example S1, into a powder concentrate A (inlet temperature 105°C; exit temperature 55°C).

15 The powder concentrate A is further dried at room temperature for 2 hours in a nitrogen-operated fluidized bed, powder concentrate B being obtained.

Characterizations:

20 Ready-to-spray mixture: 29% S.C., $7.30 \cdot 10^{11}$ cfu/g of dry matter

Powder concentrate B: water activity $a_w = 0.139$,
moisture content 3.7%,

T_g from DSC measurement: 45°C,
25 $5.06 \cdot 10^{11}$ cfu/g of dry matter (equivalent to 69% survival rate in the drying)

Storage study on powder

concentrate B: cfu counts at room-temperature storage in
30 containers sealed under ambient air:
 $4.8 \cdot 10^{11}$ cfu/g of dry matter (95%)
after 21 days

35 Example S6

The ready-to-spray mixtures were prepared in a similar manner to Example S3. Here, however, two different centrifuged ferments were used:

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Example S6a: batch fermentation, with the ferment having been cooled to 4°C for 40 minutes toward the end of the fermentation.

45 The powder concentrate A obtained in the spray-drying in accordance with Example S1 (inlet temperature from 107 to 111°C; exit temperature from 58 to 61°C) was not further dried.

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Characterizations:

Ready-to-spray mixture: $3.68 \cdot 10^{11}$ cfu/g of dry matter

Powder concentrate A: $0.76 \cdot 10^{11}$ cfu/g of dry matter (equivalent
to 21% survival rate in the drying)

Example S6b: batch fermentation, with the ferment having been heated to 44°C toward the end of the fermentation. In this example, the ready-to-spray mixture was divided. In a first experiment, the reservoir vessel was thermostatted to 4°C, as in Examples S1 to S5 and S6a. In a second experiment, the reservoir vessel was thermostatted to 20°C.

The powder concentrates A obtained by spray drying in accordance with Example S1 (inlet temperature from 103 to 110°C; exit temperature from 59 to 61°C) were not post-dried.

Characterizations for reservoir at 4°C:

Ready-to-spray mixture: $3.53 \cdot 10^{11}$ cfu/g of dry matter

Powder concentrate A: $2.36 \cdot 10^{11}$ cfu/g of dry matter (equivalent to 67% survival rate in the drying)

Characterizations for reservoir at 20°C:

Ready-to-spray mixtures: $3.53 \cdot 10^{11}$ cfu/g of dry matter

Powder concentrate A: $1.48 \cdot 10^{11}$ cfu/g of dry matter (equivalent to 42% survival rate in the drying)

Formulation examples

In accordance with the formulas stated below, dry mixtures of powder concentrates according to the invention were prepared and processed to form compacted starter culture preparations:

Unless specified otherwise, the release agent used was Leucine and the free-flowing agent used was Sipernat 50S (spray-dried silicon dioxide).

The individual components of the preparations are first mixed with one another. For this purpose, for example, a plowshare mixer is used (type Lö 20 from Lödige). The dry mixture obtained in this manner is compacted in a compactor. For example, for this purpose a laboratory compactor can be used which applies a

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pressing force of 14 kN/cm² (e.g. laboratory compactor L 200 from Bepex). The product ribbon exiting from the compactor is then comminuted to a particle size of ≤ 1.25 mm. The crude granules are screened to separate off fines of a particle size of ≤ 0.3 mm. The
5 yield of useful material is about from 50 to 60% of the material used.

Example F1: Preparing a compacted effervescent product
for use as starter culture for silage

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Preparation A:

Powder concentrate (in accordance with Example S2)	200.0 g
Citric acid, anhydrous	95.0 g
15 NaHCO ₃	95.0 g
PEG (M _w < 400)	8.0 g
Free-flowing agent	2.0 g

20 Preparation B:

Powder concentrate (according to Example S2)	100.0 g
Ascorbic acid, powder	47.5 g
NaHCO ₃	47.5 g
25 PEG (M _w < 400)	4.0 g
Free-flowing agent	1.0 g

Preparation C:

30 Powder concentrate (according to Example S2)	100.0 g
Malic acid	47.5 g
NaHCO ₃	47.5 g
PEG (M _w < 400)	4.0 g
35 Free-flowing agent	1.0 g

Preparation D:

Powder concentrate (in accordance with Example S2)	100.0 g
40 Zeolite A (Wessalith P)	20.0 g
Ascorbic acid, powder	37.0 g
NaHCO ₃	36.8 g
Release agent	3.0 g
45 Free-flowing agent	3.0 g

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Example F2: Preparation of a quick-dissolving
compacted mixture without effervescent additive

	Powder concentrate (according to Example S2)	100.0 g
5	Water-soluble surfactant (Pluriol EL 500)	90.0 g
	Release agent	7.0 g
	Free-flowing agent	3.0 g

10 Example F3: Preparation of a compacted mixture

	Powder concentrate (in accordance with Example S5)	100.0 g
	Compacting aid ¹⁾	90.0 g
15	Release agent	7.0 g
	Free-flowing agent	3.0 g

1) selected from: Avicel PH 102, Vivapur 105, FlowLac, Maltex
20, Cellactose or mixtures thereof

20 Example F4: Preparation of stabilized compacted mixtures

Base formula:

25	Powder concentrate (according to Example S5)	100.0 g
	Compacting aid	50.0 g
	Stabilizer	cf. Table I
30	Release agent	7.0 g
	Free-flowing agent	3.0 g

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Table I

	Stabilizer	Component	Amount (g)
5	A	Zeolite A	40
	B	PEG 4000	40
	C	Ascorbic acid ¹⁾	40
	D	PEG 4000	20
10	E	Ascorbic acid	20
		Zeolite A	20
	F	Ascorbic acid	20
		Zeolite A	3
15	G	PEG 4000	17
		Ascorbic acid	10
		Zeolite A	1.5
	H	PEG 4000	8.5
		Ascorbic acid	7
		Zeolite A	1
		PEG 4000	6

1) in each case L-ascorbic acid

We claim:

- 5 1. A dry microorganism culture which comprises at least one microorganism species in carrier-bound form, wherein the culture is present in the form of particles which
 - 10 a) have a particle size of at least about 0.1 mm and
 - b) are compressed.
- 15 2. A microorganism culture as claimed in claim 1, wherein the compressed particles comprise compacted broken material having a diameter of from about 0.1 mm to about 2 mm.
- 20 3. A microorganism culture as claimed in claim 1, wherein the compressed particles comprise tablets having a diameter of from about 2 to 50 mm and a ratio of diameter to thickness of from about 1:0.1 to about 10:1.
- 25 4. A microorganism culture as claimed in one of the preceding claims, wherein it comprises, as further component, an effervescent additive.
- 30 5. A microorganism culture as claimed in one of the preceding claims, wherein, as carrier, it comprises at least one matrix material for embedding the microorganism cells with or without at least one further cell-stabilizing additive.
- 35 6. A microorganism culture as claimed in one of the preceding claims, wherein it comprises from about 10^8 to 10^{12} cfu/g of at least one microorganism species.
- 40 7. A microorganism culture as claimed in one of the preceding claims, wherein it comprises at least one lactic-acid-producing bacterial species.
- 45 8. A microorganism culture as claimed in claim 7, wherein the bacterial species is selected from bacteria of the genus *Lactobacillus* sp.

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9. A process for producing a dry microorganism culture, comprising at least one microorganism species in carrier-bound form, which comprises
- 5 a) dissolving or suspending at least one substance suitable for forming a carrier in a liquid comprising at least one microorganism species,
- 10 b) drying the resultant mixture in a spray-dryer, for the spray-drying use being made of a conditioned dried gas heated to a temperature in the range of above about 80°C, and
- c) removing the dried material from the spray dryer, this dried material having an exit temperature of from about 45 to 75°C.
- 15 10. A process as claimed in claim 9, wherein, the dried gas used in stage b) has a dew point of less than about +5°C.
- 20 11. A process as claimed in claim 9 or 10, wherein, in a further stage d), the dry material is subjected to a further drying at a temperature in the range from about 15 to 50°C in a gas atmosphere or in vacuo and/or at least one desiccant is added.
- 25 12. A process as claimed in one of claims 9 to 11, wherein, as dry material, a powder concentrate having a content of viable microorganisms of from about $5 \cdot 10^8$ to $1 \cdot 10^{12}$ cfu/g is obtained.
- 30 13. A process for preparing a dry microorganism culture as claimed in one of claims 1 to 8, which comprises
- 35 i) producing a powder concentrate of the microorganism culture by carrier-bound spray-drying, carrier-bound freeze-drying or carrier-bound fluidized-bed drying,
- ii) with or without admixing the powder concentrate with one or
- 40 more coformulants and
- iii) compacting or tableting this mixture.
14. A process as claimed in claim 13, wherein the compacted powder concentrate from stage iii) is broken, with or without classification.
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15. A process for preparing a dry agglomerated microorganism culture, which comprises

5 i) preparing a powder concentrate of the microorganism culture by carrier-bound spray-drying, carrier-bound freeze drying or carrier-bound fluidized-bed drying,

10 ii) with or without admixing the powder concentrate with one or more coformulants and

iii) agglomerating this mixture.

16. A process as claimed in claim 13 or 15, wherein the spray-drying is performed as in one of claims 9 to 12.

17. The use of a microorganism culture as claimed in one of claims 1 to 8 or prepared by one of claims 9 to 16 as starter culture for foodstuffs and feedstuffs.

18. A foodstuff or feedstuff obtainable by using a microorganism culture as claimed in one of claims 1 to 8 or prepared by one of claims 9 to 16 as starter.

58/iT

Dry microorganism cultures and their preparation

Abstract

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Dry microorganism cultures comprising at least one microorganism species in carrier-bound form are present in the form of particles which a) have a particle size of at least about 0.1 mm and b) are compressed; processes for preparing dry microorganism

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cultures and their use for preparing foodstuffs and feedstuffs are also claimed.

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0050/48996

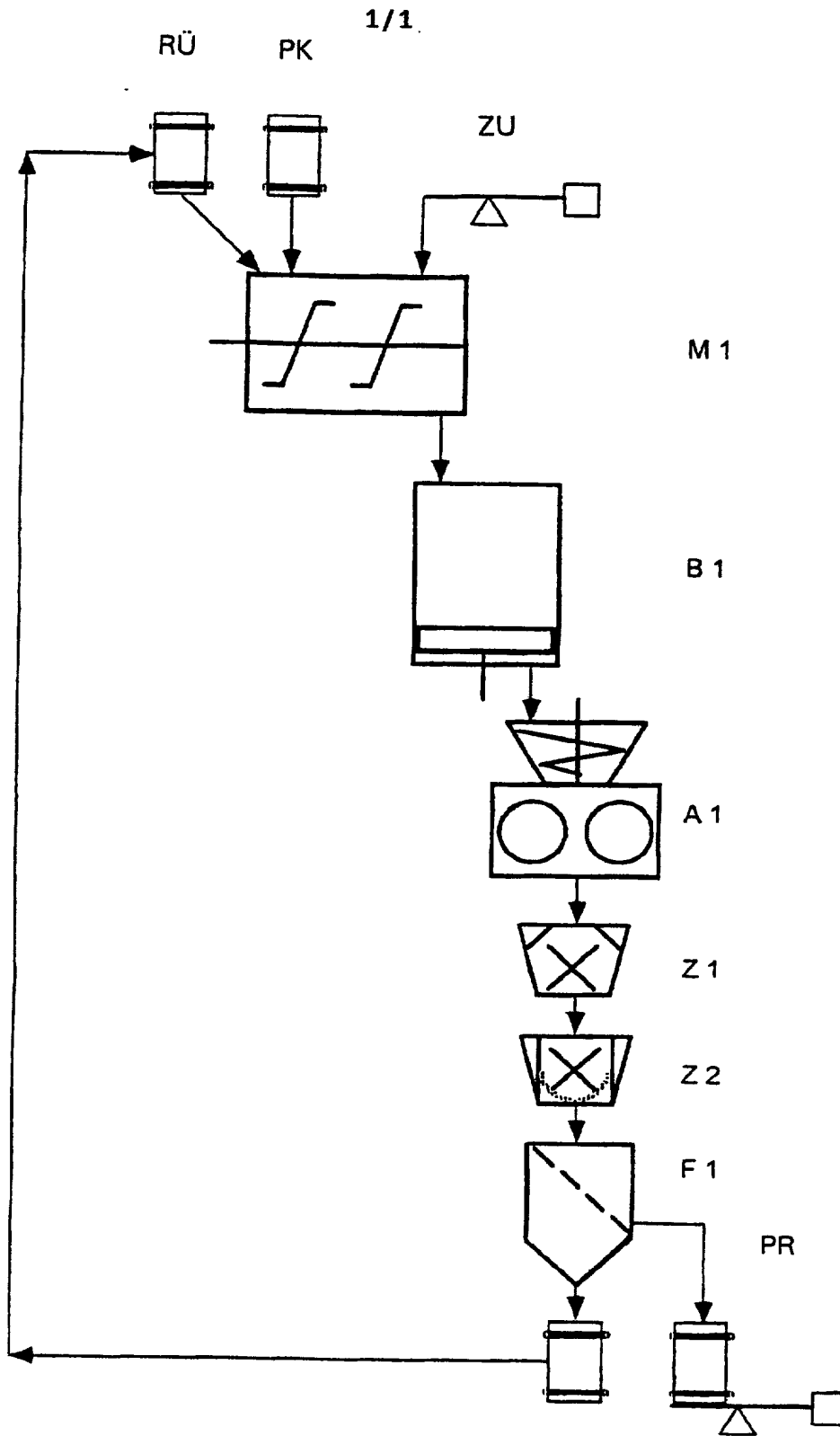


Fig. 1

Declaration, Power of Attorney

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0050/048996

We (I), the undersigned inventor(s), hereby declare(s) that:

My residence, post office address and citizenship are as stated below next to my name,

We (I) believe that we are (I am) the original, first, and joint (sole) inventor(s) of the subject matter which is claimed and for which a patent is sought on the invention entitled

DRY MICROORGANISM CULTURES AND THEIR PREPARATION

the specification of which

☒ is attached hereto.

☐ was filed on _____ as

Application Serial No. _____

and amended on _____.

☒ was filed as PCT international application

Number PCT/EP99/02925

on April 29, 1999

and was amended under PCT Article 19

on _____ (if applicable).

We (I) hereby state that we (I) have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

We (I) acknowledge the duty to disclose information known to be material to the patentability of this application as defined in Section 1.56 of Title 37 Code of Federal Regulations.

We (I) hereby claim foreign priority benefits under 35 U.S.C. § 119(a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or § 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or PCT International application having a filing date before that of the application on which priority is claimed. Prior Foreign Application(s)

Application No.	Country	Day/Month/Year	Priority Claimed
19819475.7	Germany	30 April 1998	<input checked="" type="checkbox"/> Yes <input type="checkbox"/> No

pdfiskpc001 - 25

We (I) hereby claim the benefit under Title 35, United States Codes, § 119(e) of any United States provisional application(s) listed below.

(Application Number)

(Filing Date)

(Application Number)

(Filing Date)

We (I) hereby claim the benefit under 35 U.S.C. § 120 of any United States application(s), or § 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C. § 112, I acknowledge the duty to disclose information which is material to patentability as defined in 37 CFR § 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application.

Application Serial No.

Filing Date

**Status (pending, patented,
abandoned)**

_____	_____	_____
_____	_____	_____
_____	_____	_____
_____	_____	_____

And we (I) hereby appoint **Messrs. HERBERT. B. KEIL**, Registration Number 18,967; and **RUSSEL E. WEINKAUF**, Registration Number 18,495; the address of both being Messrs. Keil & Weinkauff, 1101 Connecticut Ave., N.W., Washington, D.C. 20036 (telephone 202-659-0100), our attorneys, with full power of substitution and revocation, to prosecute this application, to make alterations and amendments therein, to sign the drawings, to receive the patent, and to transact all business in the Patent Office connected therewith.

We (I) declare that all statements made herein of our (my) own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Declaration

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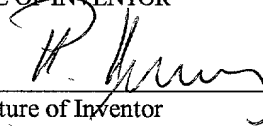
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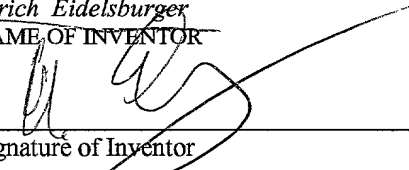
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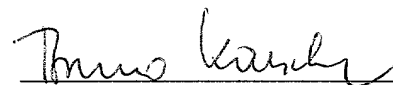
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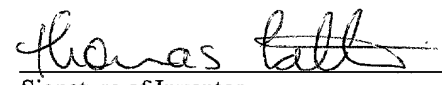
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